Review paper

Consequences of angiogenesis for tumor progression, metastasis and cancer therapy

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The growth of solid tumors to a clinically relevant size is dependent upon an adequate blood supply.1 This is achieved by the process of tumor stroma generation where the formation of new capillaries is a central event.^{1,2} Progressive recruitment of blood vessels to the tumor site and reciprocal support of tumor expansion by the resulting neovasculature are thought to result in a self-perpetuating loop helping to drive the growth of solid tumors.3 The development of new vasculature also allows an 'evacuation route' for metastatically-competent tumor cells, enabling them to depart from the primary site and colonize initially unaffected organs.4 Several molecular and cellular mechanisms have been identified by which tumor parenchyma may exert its angiogenic effect on host endothelial cells. $^{1-3,5-7}$ As a result of this paracrine influence, tumor-associated endothelial cells acquire an 'immature' phenotype¹ manifested by rapid proliferation, migration, release of proteases and expression of cytokines, endothelial-specific tyrosine kinases (e.g. fik-1, tek and others) as well as numerous other molecular alterations.3 Consequently a network of structurally and functionally aberrant blood vessels is formed within the tumor mass.8 There is also evidence that endothelial cells themselves, and likewise other stromal cells, may act reciprocally to alter the behavior of adjacent tumor cells in a paracrine or cell contact mediated fashion.3 For example, production of interleukin 6(IL-6) by endothelial cells may have a differential effect on human melanoma cells expressing different degrees of aggressiveness.9 in this manner endothelial derived cytokines could conceivably contribute to tumor progression by suppressing the growth of the less aggressive tumor cells and promoting dominance of their malignant counterparts in 'strategic' perivascular zones. Distinct biological features expressed by tumor-associated vasculature may serve as potential prognostic markers of

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disease progression as well as novel targets for therapeutic intervention.

Key words: Angiogenesis, cancer therapy, metastasis, tumor progression.

Introduction

The rapid progress made in molecular methodology over the last two decades has resulted in an impressive series of discoveries regarding the genetic events involved in tumor development and progression. Activation of dominantly acting oncogenes or loss of recessively acting tumor- or metastasis-suppressor genes have been frequently detected in various types of cancer. 10,11 These remarkable advances in our understanding of the nature of malignant transformation have promoted the reductionist notion that cancer—an otherwise multicellular and extremely complex disease—can be understood primarily if not exclusively on the basis of a detailed analysis of the growth control abnormalities expressed by various clonal tumor cell lines in tissue culture.

Until recently most models describing and explaining the course of tumor progression focused on aspects of tumor cell autonomy, and put much less emphasis on the role of tumor-host interactions as an essential part of the process. ^{11,12} Recent findings, however, suggest that several genes, although fulfilling criteria of tumor or metastasis suppressors, do not necessarily affect tumor cell proliferation directly. Rather their activity seems to be necessary to restrain tumor growth promoting cell-cell or cell-extracellular matrix interactions. This mode of action has been suggested for several different molecules including tissue inhibitor of metallo-proteinases (TIMP), ¹³ E-cadherin, ¹⁴ DCC, ¹¹ connexin 32, ¹⁵ a suppressor gene that controls thrombo-

spondin expression¹⁶ and a number of other molecules. ^{17,18}

Studies on the role of oncogenes and tumor suppressor genes in the pathology of tumor stroma generation are clearly gaining momentum in current efforts to understand basic principles of tumor growth and metastasis.⁵ For example, loss of a tumor suppressor gene controlling the oncogenic behavior of BHK cells results in conversion to a tumorigenic phenotype and is strictly coordinated with a shut off in the expression of an angiogenesis inhibitor homologous to thrombospondin (TSP). Similarly in fibroblasts from patients with Li-Fraumeni (L-F) syndrome, spontaneous loss of the remaining single copy of the wild-type p53 tumor suppressor gene leads to a down-regulation of TSP expression, 19 supporting the notion that p53 regulates TSP, which in turn regulates angiogenesis and endothelial cell growth. Interestingly p53 gene 'knock-out' in mice leads to a high frequency (30%) of endothelial neoplasia. 20 In another study, transgenic mice carrying the neu oncogene driven by an MMTV promoter were found to develop mammary tumors, which were not only highly aggressive, but also highly angiogenic.²¹ (Muller, personal communication.) It has also been reported that progression of benign hyperplasia to rapidly growing fibrosarcomas in mice carrying an oncogenic transgene is associated with the onset of secretion of the highly angiogenic cytokine, basic fibroblast growth factor (bFGF).²² Similarly, in isolated pancreatic islets from transgenic mice carrying a viral oncogene driven by the insulin gene promoter, the acquisition of angiogenic potential preceded tumorigenic conversion by these pancreatic multicellular aggregates.23 Some of the generic mechanisms found to be involved in the 'angiogenic switch' during tumor progression are listed in Table 1. Overall, such findings are consistent with the view that the nature of the growth control defect in cancer comprises molecular mechanisms operating within transformed tumor cells but also includes intercellular interactions at the interface between tumor and host.³

Reciprocal interactions between tumor cells and host stromal cells, in addition to affecting tumor growth, ^{24,25} may also contribute to tumor cell invasion of the surrounding tissues and metastatic dissemination to distant organs. In this respect, recent evidence suggests that some of the proteolytic enzymes generally associated with tumor cell invasion and metastasis are in fact produced by 'activated' stromal cells rather than by tumor cells *per se*. These stromal cell associated enzymes include, e.g. stromyelysin 3, ²⁶ collagenase type IV²⁷ and plasminogen activator (uPA).²⁸

Thus, interactions between transformed parenchymal tumor cells and 'activated' host stromal cells are likely to contribute to a self-accelerating chain of events driving tumor progression as well as tumor growth.³ Consequently, therapeutic strategies aimed at disrupting such tumor-host interactions could facilitate stabilization of the disease and possibly lead to improvements in long-term survival rates, if not cure. In this regard targeting hostderived blood vessels within solid tumors offers a unique therapeutic opportunity based on the absolute dependence of solid tumor growth on this single stromal component.²⁹ The remainder of this review will focus on some of the interactions between tumor cells and endothelial cells as well as the consequences these interactions may have on tumor behavior and success or failure of anti-cancer therapy.

Table 1. Molecular mechanisms of the 'angiogenic switch'

| Generic mechanism | Molecule involved (example) | Tumor type | Observation | Reference |
|---|--|--|---|--|
| Onset of TAFa | ? | pancreatic islets in | transformation is | 23 |
| production | bFGF | transgenic mice; hu- man melanoma | associated with the onset of TAF production | 96 |
| Onset of TAF secretion | bFGF | fibrosarcoma in mice carrying BPV trans gene | onset of the bFGF export during tumor progression | 22 |
| Loss of angio- genesis inhibitor expression | thrombospondin, (p53) | BHK cells; human breast cancer cells; trans formed L-F fibroblasts | loss of tumor suppressor gene is associated with loss of TSP expression | 19, 154 |
| 'Masterswitch ef- fect' | ras; genes regu- lated by ras (TGFa, VPF?) | intestinal cancer; colon cancer? | ras transformed cells acquired angiogenic phenotype | J Rak, J Filmus and RS Kerbel, unpublished |

^aTAF, tumor angiogenesis factor.

Tumor progression and metastasis

The fundamental observation that tumor growth is not only associated with an increase in size of the lesion but, more importantly, with qualitative changes in its characteristics, led to the concept of 'tumor progression'. 30 It is generally believed that during this process tumor cells acquire features of greater malignancy. Some examples include hypersensitivity to growth factors resulting in increased growth potential, invasiveness, metastatic competence as well as resistance to normal regulatory cytokines and therapeutic agents. 30-32 The pace of these changes varies between individual cases, ranging from slow and continuous to stepwise, rapid and sometimes fulminant. For example, rapid, probably onestep acquisition of a highly malignant phenotype is observed in what is referred to as 'type II progression'. This type of progression is frequently linked with cases of so-called 'unknown primary tumors' (UPT).³³ By definition, the first manifestation of UTP is an advanced and usually deadly metastatic disease, without the preceding development of an apparent primary lesion.³³ Another aspect that may vary is the pathway of progression of a particular tumor, i.e. the type, seqence and final outcome of changes observed in the phenotype of transformed cells over time. 30,34,35

It has been postulated that multiple genetic changes observed during human tumor progression may result from either activation of a 'master switch' gene followed by coordinated expression of a particular set of 'responder' genes³⁶ or separate genetic alterations may occur independently.¹¹ In this regard a breakthrough came from studies on genetic correlates of colon carcinoma progression.¹¹ Studies by Vogelstein and colleagues identified sequential chromosomal and genetic abnormalities accompanying clinical steps of disease progression.11 The sequence with which all these alterations occur is not inviolate and it appears to be their accumulation rather than order which is of biological significance.11 Similar studies have also been conducted on many other human malignancies, including glioblastoma and melanoma. 37,38

The discovery of cumulative genetic alterations in colon carcinoma and other tumor types in conjunction with the fact that the vast majority of tumors were found to be clonal in origin³⁹ understandably highlighted the 'cellular' dimension of the tumor progression process. This is consistent with the notion that the process can essentially be described as a 'clonal evolution' of the transformed cellular lineage.^{12,31} According to this concept, the genetic

instability of tumor cells and selection by the microenvironment constitute the two main forces driving this cellular 'evolution'. Consequently several cycles of tumor cell diversification and selection could lead to the emergence of a rare 'species', a highly malignant cellular variant, capable of invading the surrounding tissue and metastasizing to distant organs. The execution of the metastatic program would therefore naturally require the expression of a highly specialized phenotype in order to avoid destruction while traversing the many steps and physiological barriers of the 'metastatic cascade'.40 For example, tumor cells would have to leave the primary site, survive the highly traumatic voyage in the blood stream, extravasate and then expand in the foreign tissue microenvironment of various distant organ sites. 40,41 As predicted by the 'decathlon champion' model of metastasis, even a small minority (a clone) of metastatic cells could eventually kill the host by virtue of their high efficiency in colonizing vital organs outside the primary tumor.40 Indeed highly metastatic variants have been isolated from various animal and human tumor cell lines or surgical specimens using sequential in vivo selection protocols. 32,40 Based on these findings it was anticipated that cells isolated from primary tumors should express entirely different phenotypes than cells that are already present in the metastatic foci. Somewhat surprisingly this was infrequently found to be the case.³² On the contrary, cells representative of advanced primary tumors and their metastases were often found to be similar or even identical with respect to multiple features, including metastatic competence in experimental animals. 32 This is now thought to be a result of the fact that along with the other necessary components required for metastatic competence, tumor cels are co-selected for progressive growth properties which endow the metastatic subpopulation with a selective growth advantage. The latter feature may eventually lead to partial or complete 'clonal dominance' of metastatically competent tumor cells at the primary tumor site.³² Quantitative enrichment in those aggressive cellular variants is likely to increase the probability of invasion and formation of secondary tumor deposits in distant organs.^{3,32} Thus in addition to intracellular genetic changes, the process of tumor progression also appears to rely on changes occurring at the whole cell population level. Again, several studies have pointed out the fact that many properties of tumors, including growth rate, drug resistance, metastatic capacity and survival, cannot always be predicted by a simple analysis of separated tumor cell clones. Rather, formation of

multicellular functional 'units', 'community effects' or other types of cellular interactions operating in this cellular 'society' are believed to be highly relevant for the expression of the respective tumor properties. ^{42–48} The recruitment of host stromal cells in general, and angiogenesis in particular, add another level of complexity to the cellular interactions accompanying tumor progression.

Tumor angiogenesis

Solid tumor growth in vivo beyond 1-2 mm in diameter is almost without exception associated with recruitment of new blood vessels. 1,29,49 This is a consequence of the release of angiogenic factors by tumor cells themselves or by infiltrating inflammatory cells. 50,51 There are a number of angiogenic factors which are known to induce the process of new blood vessel formation. 1,3,6,7 Resulting angiogenesis is a multistep process that involves sprouting and subsequent capillary loop formation of endothelial cells, most likely originating from local post-capillary venules. 1,6 The angiogenic stimulus is believed to trigger several functional responses in endothelial cells, including local basement membrane dissolution, endothelial cell migration, proliferation and microvessel morphogenesis. 1,6 The process appears to be regulated by a number of polypeptide cytokines, but also by fibrin, various extracellular matrix molecules, integrins and other adhesion molecules, 52,53 proteolytic enzymes, 54,55 various carbohydrates including hyaluronan,56 carbohydrate binding proteins, 57,58 steroid hormones, 59,60 lipids, metal ions, and a variety of different small molecular weight substances.61 An updated list of angiogenic factors and molecules identified thus far can be found in some of the review articles recently published on the subject. 1,5-7,62 Alternative mechanisms of blood vessel formation which do not involve sprouting have also been described recently. 63,64

In order to measure the angiogenic activity of tumor cells and their angiogenic factors, a number of angiogenesis assays have been developed. Efforts to use these assays as a means of accurately quantitating angiogenic activity as a function of tumor type, disease progression or response to therapy have been largely unsuccessful. It is conceivable that difficulties in quantitating the stimulatory ('afferent') component of the angiogenic reaction may be due to the fact that the response of endothelial cells to angiogenic cytokines is not always linear. For example, endothelial cells respond in a

somewhat biphasic fashion to increasing concentrations of angiogenic polypeptides such as acidic or basic fibroblast growth factor (aFGF and bFGF) or tumor necrosis factor (TNF)-a, both in vivo and in vitro^{66,67} (Rak, unpublished observations). Interestingly, a number of angiogenesis stimulators, including bFGF, are produced by endothelial cells themselves, either constitutively or upon stimulation, and may act on these cells in an autocrine manner. 68,69 The latter situation may conceivably lead to an 'explosive' self-amplifying event rather than a typical dose-dependent process limited by the availability of the stimulant. On the other hand, the aforementioned angiogenesis assays have been used successfully for quantitation of angiogenesis inhibitors. 66 This may be explained by the fact that angiosuppressive agents may interfere predominantly with the rate limiting, effector ('efferent') arm of the angiogenic process.

The course of angiogenesis probably depends on multiple factors including the nature of the inducing stimulus and characteristics of local endothelial cells in a particular target organ. Nevertheless, there seem to be two generic differences between angiogenesis taking place in normal as opposed to tumor tissue. First, tumor expansion is continuous and this usually triggers chronic vascular growth and remodeling rather than finite repair processes.^{2,8,70} Second, the tumor neovascularization leads to a wide range of structural and functional blood vessel abnormalities, including tortuosities, dilatations, arterial-venous (A-V) shunts and trifurcations,8 that are normally not found during the course of 'normal' angiogenesis. In addition, tumor-associated blood vessels also have poorly developed basement membranes and lack collaterals, innervation and pericytes. Some of these factors may contribute to blood vessel leakiness. Blood vessel permeability may also be regulated by certain cytokines such as the angiogenic polypeptide vascular permeability factor (VPF), also known as vascular endothelial growth factor (VEGF), 71,72 which is secreted by various types of

These multiple abnormalities in blood vessel function and architecture are likely to result in disturbances in blood flow, with frequent stasis and changes in flow direction, leading to impairment in metabolite exchange. ^{8,75} In the absence of functional lymphatic drainage of the tumor tissue, the resulting high interstitial pressure within solid tumors may further compromise the penetration of nutrients into the deeper layers of the tumor and ultimately lead to local necrosis. ⁸ Thus tumor perfusion

appears to be inefficient, despite an abundant and continuously expanding neovasculature.

Interactions between heterogenous tumor cell subpopulations and the vasculature

As discussed at the outset, it has been well documented that even within a single tumor lesion tumor cells are highly heterogenous. 40,44,76 It is also believed that continuous generation of new cellular variants leads to the acquisition of more invasive, metastatic or drug resistant properties.32,76 Consequently, various tumor cell subpopulations may also differ in their ability to elicit neovascularization.3,44,77-79 For example, different types of angiogenic signals may be released by different subpopulations of tumor cells due to variable expression of a number of genes encoding potentially angiogenic cytokines. Some of these 'tumor angiogenesis factors' may be expressed constitutively or may be switched on in response to local microenvironmental stimuli. For example, recent studies suggest that under hypoxic conditions, glioblastoma cells switch on expression of VEGF/VPF, which otherwise is usually not expressed by this tumor type. 73,74 It is also possible that angiogenic signals may be generated by host stromal or inflammatory cells present at the tumor site as a result of stimulation by paracrine factors elaborated by various subpopulations of tumor cells. 3,21,50,80 Conversely, as will be discussed later, some endothelial cell pro-

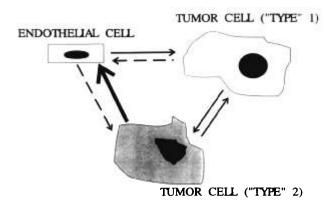


Figure 1. Interactions of heterogenous tumor cells with endothelial cells. Endothelial cells not only receive angiogenic stimuli but may also deliver regulatory signals to tumor cells. Tumor cell subpopulations (types) may differ in their capacity to generate and respond to those regulatory signals. Interactions between heterogenous tumor cells and endothelium are likely to be: 1, reciprocal; 2, dynamic; 3, selective; 4, direct and indirect.

ducts may also selectively alter the behavior of adjacent tumor cells. Thus tumor-endothelial cell interactions may generally be thought of as reciprocal, dynamic and selective (Figure 1).

Tumor cell subpopulations are likely to emerge and expand within the tumor mass in certain microdomains that would require a distinct blood vessel supply for their continued growth. If tumor cell subpopulations occupying adjacent microdomains differ in their angiogenic capacity one would anticipate finding greater blood vessel densities in some microregions than in others.3 Indeed such vascular 'hot spots' have been found in tissue specimens from breast and prostate cancer.81-84 More importantly, the dense vascularity of these 'hot spots' appears to correlate with an unfavorable prognosis in these tumor types, 81 suggesting that tumor cells occupying these regions are likely to be both highly angiogenic and metastatic.³ It is also possible that blood vessels attracted by a more angiogenic tumor cell subpopulation may facilitate growth, and even metastasis, of adjacent and relatively less angiogenic tumor cells. 44,77 Although demonstrated in some cases,⁷⁷ this 'co-operative mode' of tumorendothelial cell interaction stands in contrast to work which has shown that more malignant (and presumably more angiogenic) tumor cells sometimes overgrow ('dominate') their less aggressive counterparts in vivo.32 We postulate therefore that more often than not, different tumor cell subpopulations 'compete' rather than 'co-operate' during tumor neovascularization and that a greater angiogenic capacity may endow tumor cells with a selective growth advantage³ (Figure 2).

The role of angiogenesis during tumor progression: cutaneous malignant melanoma as a paradigm

Blood vessels recruited by angiogenic stimuli contribute to solid tumor growth by maintaining perfusion and hence viability of tumor parenchyma. During tumor development the 'switch to the angiogenic phenotype' was shown to be strictly correlated with the acquisition of progressive growth properties by the tumor cells. Purple Furthermore, the increase in blood vessel counts in hypervascular 'hot spots' was shown to be associated with a poor prognosis in breast or prostate cancers. Vascularity has also been associated with tumor progression in cutaneous melanoma. 35,49,78,86–89

Cutaneous malignant melanoma progression recently attracted much experimental attention for a

1.COOPERATIVE MODE (MILLER 1990, JOUANNEAU 1994)

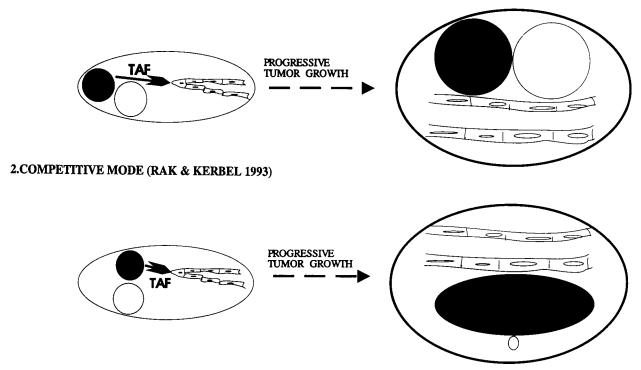


Figure 2. Two current hypotheses predicting that either the competitive or the co-operative mode of interaction governs tumor subpopulation expansion. Tumor cells producing tumor angiogenesis factor (TAF) may either facilitate expansion of the adjacent less angiogenic counterparts or successfully compete with the latter for blood vessels and thereby acquire a selective growth advantage.

number of reasons. Poor prognosis and rapid development of clinically malignant disease were found to correlate with increasing thickness of primary lesions, especially beyond a threshold level of 0.76 mm. 90-92 Because lesions from different phases of tumor development are readily accessible for observation and sampling, it became possible to describe distinct stages of melanoma progression and sometimes establish corresponding cell lines in tissue culture. 91-93 It is believed that early, curable melanoma usually begins as a so-called radial growth phase (RGP) lesion which is thin, often confined to the epidermis and avascular. Expansion of melanoma into the dermal mesenchyme is associated with the vertical growth phase (VGP) of the tumor. During this phase the lesion is evolving towards an invasive, thick, vascular and eventually incurable metastatic tumor. 89,91,92,94,95

Melanoma cell lines representative of sequential stages of tumor progression express a host of differential characteristics reminiscent of their in situ phenotype. Differences in production of various cytokines, some of which possess angiogenic activity, have been noted between cells from 'early' versus 'late' melanoma lesions. 78,96,97 Differences

also exist between early and late stage melanoma cells in their responsiveness to a number of growth inhibitory cytokines including TGF-\(\beta\), IL-1, IL-6, OSM and others. Progression of early phase melanoma cells to a state of so-called 'multicytokine resistance' appears to parallel the increase in clinical aggressiveness of the respective lesions. 94,98-101 Interestingly, endothelial cells are also capable of producing a number of these potentially growth inhibitory cytokines.^{3,9} In fact, it has recently been documented that during angiogenesis, endothelial cells overexpress at least one of these cytokines, i.e. IL-6. 102 It is also known that, although RGP melanomas usually remain avascular, the tumor cells, even at this early phase of disease progression, produce some potentially angiogenic polypeptides, including bFGF. 96,97,103 One would anticipate then, that once these angiogenic factors are released, the resulting onset of angiogenesis in an early stage melanoma may actually have a somewhat 'suicidal' effect on the tumor cells themselves. in that it would expose these cytokine-sensitive cells to endothelium derived growth inhibitors.³ For further tumor growth it would seem necessary that this effect be overcome by selection of cytokine

resistant tumor cell variants. Indeed, we have recently demonstrated that melanoma cells isolated from different stages of the disease progressively lose their sensitivity to endothelial cell derived growth inhibitors, including IL-6.9 That is consistent with results reported recently by Barnhill et al. 89 These investigators found that transition from RGP to VGP melanoma is frequently associated with a sudden burst of angiogenesis. Paradoxically the high blood vessel count of the 'progressive' RGP tumors coincided with histological 'regression' of the tumor and with a greater risk of metastasis at the later stages of disease progression. 89,104 The most recent evidence indicates that high vascular counts detected in thin primary melanomas, which otherwise carry a favorable prognosis, is predictive of recurrent, fatal metastatic disease. 105 Such a predictive association was also observed in thicker, more advanced primary lesions by some investigators 49,86-88 but not by others. 105,106 Thus a causal

relationship may exist between the initial 'switch to the angiogenic state' and histological regression followed by progression of RGP melanoma toward metastatic disease (Figure 3). Tumor progression in this case may be driven by selection of more malignant cellular variants that can withstand the exposure to inhibitory cytokines associated with the presence of an abundant vasculature, whereas their cytokine-sensitive counterparts may undergo complete or partial extinction. J. 107, 108 It is not clear whether the regression observed in cases of hypervascular RGP melanoma can be entirely attributed to endothelial derived IL-6 or if other inflammatory cytokines are involved as well.

Also of interest is the fact that IL-6 was shown to stimulate the motility of breast cancer cells *in vitro*. ¹⁰⁹ It is tempting to speculate that in breast cancer greater motility (invasiveness) of cancer cells may be induced by the presence of adjacent endothelial cells in vascular 'hot spots' as observed in

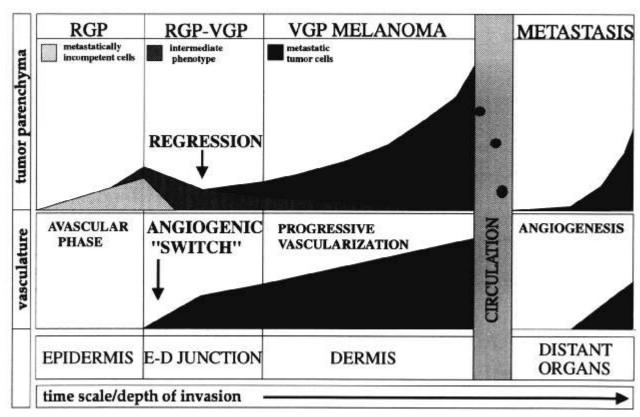


Figure 3. Interrelationship between angiogenesis and the stage of progression in malignant melanoma. Transition from RGP to VGP is frequently associated with histological regression within the lesion. This regression coincides with a 'switch' to the highly angiogenic state. At this stage of tumor progression abundant blood vessels are frequently observed. In the presence of endothelial cells and inflammatory cells the more aggressive melanoma cell populations (black) may acquire a growth advantage over their non-metastatic counterparts (light gray; dark gray represents intermediate phenotypes). These effects may be mediated by IL-6 and other cytokines that can selectively inhibit the growth of early stage melanoma cells. Further increase in tumor vascularity, a secondary 'angiogenic switch', is frequently observed during the later stages of disease progression. This cascade of events may facilitate tumor cell dissemination and metastasis.

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breast cancer.⁸⁴ This may be another explanation for why a high blood vessel count in these 'hot spots' is thought to be a correlate of subsequent metastatic disease.⁸⁴

Characteristics of tumor-associated endothelial cells

Cells from the endothelial lineage express multiple levels of heterogeneity. 110 For example, even endothelial cells in the same vessel have been shown to display different turnover rates. 111 Moreover, endothelium from large vessels was shown to be phenotypically and functionally different from microvascular endothelial cells. 112 It is also known that isolated endothelial cells in tissue culture can be highly diverse in terms of their growth and tubule forming capacity, as well as other properties. 110,113,114 It may be that some kind of stem cell-like hierarchy exists in the endothelial cell lineage that allows a self renewal of these cells upon injury.3 Furthermore, organ- or vascular bed-specific features have been found in different types of endothelium. 110,115 Some of these features have been implicated in organ-specific homing of lymphocytes. 116 In addition, organ preference can apparently play an important role in the phenomenon of organ-specific metastasis. 117 The differential expression of adhesion molecules and growth or motility factors by endothelial cells in different vascular beds was implicated in this so called 'seed and soil effect'. 117,118

Although it is conceivable that certain properties of tumor-associated endothelial cells are to some extent site-dependent, ¹¹⁵ there is also evidence that distinct phenotypic features are induced by the tumor microenvironment itself. Recent animal studies have shown that the labeling index of tumor-associated endothelial cells is up to 50-fold higher than their quiescent counterparts from healthy tissues. ¹¹⁹ This high proliferation rate is likely to be due to the imbalance between angiogenesis stimulators and inhibitors at the tumor site. ⁵⁴

At the molecular level, the 'activated' (and functionally immature) endothelial cell phenotype is associated with a unique pattern of gene expression. ¹²⁰ Some of the molecules expressed by tumor-associated (but not by normal quiescent) endothelial cells have been identified using monoclonal antibodies, e.g. the difference in expression of the antigen recognized by the EN7/44 monoclonal antibody. ¹²¹ This antibody was raised against highly vascular fragments of human breast carcinoma tis-

sue, and was found to preferentially react with budding blood vessels in breast cancer and mammary hyperplasia. Another antibody, designated FB5, originally raised against cultured fetal fibroblasts, was shown to react with endosialin molecules expressed specifically by blood vessels associated with several types of human cancer. Similarly, the recently described antibody 4A11 preferentially reacts with endothelial cells in adrenal tumors, Kaposi's sarcoma and a variety of inflammatory lesions. Two monoclonal antibodies called TEC4 and TEC11 were shown to recognize endoglin, a cell surface proteoglycan that is expressed selectively by tumor-associated endothelial cells. 124

A distinct pattern of gene expression in tumorassociated endothelial cells was also detected at the mRNA level. For example, overexpression of mRNA for the c-ets transcription factor (and oncogene) was detected in tumor-associated blood vessels as well as in exponentially growing or cytokine-stimulated endothelial cells in culture. 125 This gene is believed to be involved in regulating the expression of extracellular matrix degrading enzymes which are thought to play a role in angiogenesis. 125 Endothelial cell specific receptor tyrosine kinases (see Table 2) encoded by the genes tie-1, 126 flk-1 127,128 or tek¹²⁹⁻¹³¹ have been recently described. Both flk-1 and tek receptor tyrosine kinases are expressed by endothelial cell precursors during embryogenesis in the mouse^{128,129,132,133} and also in blood vessels associated with xenotransplants of human malignant melanoma cells in nude mice (Yamaguchi and Rak, 1991, unpublished). No such expression of flk-1 or tek was observed in surrounding healthy tissue in tumor-bearing mice¹²⁸ (Yamaguchi and Rak, 1991, unpublished). The similarity between 'embryonal' and 'tumor associated' endothelial cells with respect to flk-1 and tek gene expression suggests that tumor microenvironment may induce or select some sort of 'embryonal' or 'stem cell' phenotype in endothelial cells recruited to the tumor site. Since flk-1 acts as a functional receptor for vascular permeability factor VPF/VEGF, 132,134 interference with its function would be expected to produce some kind of anti-angiogenic and antitumor effect. Indeed, treatment of tumor-bearing mice with a neutralizing antibody to VEGF was found to produce such an effect. 135 Moreover co-injection of C6 mouse glioblastoma cells with cells producing a retrovirus encoding a defective mutant of the flk-1 gene resulted in a dominant-negative effect on VEGF binding capacity of the *flk-1* gene product. As a result, both angiogenesis and tumor growth were profoundly inhibited. 136 The significance of

Table 2. Endothelial RTKs

| RTC | Other designations | Ligand | Expression | References |
|-----------|-------------------------------------|----------|--|---|
| flg | <i>fms</i> -like gene, FGFR1 | bFGF | endothelial cells in culture, tumor associated endothe- lium | 156, 157 |
| bek | FGFR2 | bFGF | endothelial cells in culture | 157 |
| flk-1 | KDR | VEGF/VPF | angioblasts, tumor associated endothelial cells, kidney, brain | 73, 74, 132, 134, 137 |
| flt (1–4) | <i>fms</i> -like tyrosine kinase | VEGF | embryonal blood vessels, human fetal lung, kidney, liver | 127, 139, 141 |
| tie-1 | | ? | early embryonal vascular system | 126, 158 |
| tie-2 | | ? | angioblasts, sprouting endothelial cells, embryonal organs | 130, 131, 158 |
| tek | tie-2 truncated form | ? | embryonal blood vessels, tumor associated endothe- lial cells | 129, Yamaguchi and Rak, unpublished (1992) |

this finding stems from the fact that the expression of VPF/VEGF by tumor cells and its corresponding receptor by endothelial cells has been described in at least two other clinical and experimental tumors including glioblastoma multiforme, as well as cancers of the urinary tract and kidney. 73,74,137,138 Interestingly, proliferating endothelial cells in tissue culture overexpress another VEGF receptor tyrosine kinase (flt) the role of which is not fully understood. 139 In addition, hemangioma-like endothelial outgrowths have been observed in transgenic mice overexpressing a different receptor tyrosine kinase, flp. 140 Again, it is not known whether the latter tyrosine kinase is expressed and plays a role in solid tumor-induced angiogenesis. A potential role in this process was, however, recently implicated for tyrosine kinase receptors binding bFGF. In these experiments overexpression of the high affinity receptor transcript and bFGF receptor protein was demonstrated in blood vessels of transplantable murine tumors, (J Gross-Dzubow, personal communication). Somewhat paradoxically, however, the flg gene product, another fibroblast growth factor receptor with tyrosine kinase activity, was shown to be upregulated when endothelial cells exit the phase of rapid proliferation. 141

Altered expression of several molecules unrelated to cytokine receptors has also been recently reported in tumor-associated endothelium. For example, abnormal expression of the fibronectin receptor on the luminal surface of rapidly growing endothelial cells or MHC class II antigen expression

on interferon (IFN)- γ activated tumor associated endothelium was observed. It is noteworthy that some unique characteristics of vascular endothelium may change with tumor progression and such alterations could be explored as potential targets for anti-vascular or anti-angiogenic cancer therapy.

Targeting of tumor-associated blood vessels as a novel anti-cancer treatment strategy

Several considerations make targeting the tumor vasculature an attractive possibility as a form of anti-cancer therapy. 144-148 First, tumor cell genetic instability and resulting heterogeneity are thought to be two of the main factors causing the rapid development of tumor cell resistance to natural growth control mechanisms as well as therapeutic agents. By contrast, normal stromal cells should be genetically stable and hence more likely to remain susceptible to exogenous therapeutic influences, i.e. their capacity to develop resistance should be minimal, in comparison to tumors. 148 Second, the accessibility of endothelial cells to blood borne pharmacological manipulations makes them particularly attractive for such stromal cell targeting. Targeting the tumor vasculature is also a strategy that might circumvent the problem of limited penetration frequently encountered when delivering high molecular weight drugs to solid tumors with elevated interstitial pressures.8 Such penetration would not be necessary when targeting the vasculature. It

is thought that antivascular therapy would simultaneously obliterate the capacity of the tumor to continue to grow and metastasize. Third, the effect of vascular damage on tumor growth is likely to be amplified by the fact that the viability of numerous tumor cells depends on a much smaller number of functional capillaries. Finally, tumor-associated endothelial cells express characteristics which are not normally found in the vasculature of tissues outside the tumor, and this may allow the design of highly specific and non-toxic treatment strategies.

Possible specific strategies for executing an antivascular attack have been suggested by several investigators and have led to experiments with a variety of anti-angiogenic agents (Table 3). It would seem reasonable to attempt to obliterate the angiogenic circuit driving blood vessel formation in a particular tumor. This could be achieved by targeting the relevant angiogenic factor, its receptor or,

Table 3. Anti-angiogenic and angiosuppressive agents 144–147,159,160

Natural angiogenesis inhibitors platelet factor 4 (PF4) TIMP-1, TIMP-2 thrombospondin cartilage derived inhibitor (CDI) IFN- α and - β Angiostatin Antagonists of angiogenic growth factors suramin and 254 compound pentosan polysulfate (PPS) Angiostatic steroids and their co-inhibitors cortisone medroxyprogesterone heparin (fragments) β-cyclodextrin tetradecasufate HNT Anti-inflammatory drugs indometacin ibuprofen Natural antiangiogenic compounds and their derivatives angioinhibins (fumagilin, AGM-1470) vitamine D₃ analogs herbimycin A retinoids somatostatin analogs sulfated chitin derivatives Anti-cancer drugs flavone acetic acid (FAA) bleomycin metotrexate mitoxantrone Other copper chelators (p-penicillamine) minocycline mitotoxins (FGF-saporin)

alternatively, properties of activated endothelial cells such as growth, migration, proteolysis or morphogenesis using specific antibodies, inhibitors or receptor antagonists. 145,149,150 One difficulty in executing this 'anti-angiogenic' approach is that the exact mechanism of angiogenesis operating in any given tumor is usually unknown and multiple angiogenic factors are likely to be involved. It is conceivable that the angiogenic capacity could perhaps be 'exhausted' by interference with intracellularly acting cytokines regulating growth, survival and senescence of endothelial cells. Potential target molecules for this kind of intervention would likely include IL-1, FGF¹⁵¹ or IL-6 (Rak, unpublished).

Another strategy, originally proposed by Denekamp, is to target directly a known characteristic of tumor associated endothelial cells thought to be critical, i.e. their high proliferation rate. 146,157 This 'angiosuppressive' effect could be accomplished by using endothelial cell-specific cytotoxic agents including derivatives of known chemotherapeutic drugs. In fact some of the effects of treatment with IFN, certain cytokines, ionizing radiation, hyperthermia and even certain anti-cancer drugs may be attributable, at least in part, to blood vessel damage leading to hemorrhagic necrosis of the tumor. 146 The available drugs studied so far, however, usually lack sufficient endothelial cell specificity and can therefore cause severe toxicity. It seems somewhat paradoxical that although essentially all of the antiproliferative chemotherapeutic agents should cause some degree of vascular damage this has not been noted with any consistency suggesting that some degree of drug resistance may also be expressed (or induced) in tumor-associated endothelial cells. 148 Alternatively, specific immunotoxins, retroviruses or antisense reagents can be designed to target genes expressed by tumor-associated endothelial cells, including KDR (human homolog of *flk-1*) and other tyrosine kinases.^{3,136}

It is possible that different types of tumors will require different anti-angiogenic treatment strategies. For example, recent studies show that at least in human breast carcinoma the proliferating fraction of endothelial cells is much smaller than one would have expected based on previously cited rates most of which were obtained using animal models. ^{152,153} Thus in this disease the expansion of the microcirculation may be brought about by endothelial cell migration and remodeling rather than cell growth *per se.* ¹⁵² In such cases antiproliferative agents are likely to be ineffective and should be replaced by protease inhibitors, steroids or other compounds interfering with endothelial cell migration, base-

RGD polymers

ment membrane turnover and morphogenesis.¹ It has also been postulated that blood channels may develop in a tumor mass in areas where the endothelial lining is incomplete or absent. It is unclear how blood flow could be maintained under these highly coagulogenic conditions but certainly in such a situation specific anti-endothelial therapy would be less effective.⁶

Anti-angiogenesis therapy of solid tumors: present problems and future directions

Clearly, there are a number of appealing approaches to treating solid tumors by inhibition of tumor angiogenesis. The main problem will undoubtedly be to achieve selectivity: are there molecular targets associated with new blood vessel formation that are not found on normal, mature blood vessels? In a sense this requires shifting our traditional focus on developing 'magic bullets' specific for tumor cells to designing or discovering similar magic bullets which show selectivity for tumor-associated blood vessels. However, even if such complete selectivity cannot be attained it does not exclude—as chemotherapy and radiotherapy have shown—the possibility of using anti-angiogenesis strategies to treat solid tumors.

A second problem stems from the assumption that complete kill of solid tumor masses may never be attainable since small (e.g. 1 mm) tumors can survive without a blood vessel supply. However, antiangiogenesis may be an excellent paradigm for the concept of treating cancer by *controlling* progressive tumor growth as opposed to trying to achieve total tumor cell *kill*. This may lead to long-term disease stabilization in some cases.

So what does the future hold? Obviously there will continue to be a great emphasis on finding anti-angiogenic agents or drugs which have the desired magic bullet properties. As such drugs are uncovered, the appropriate phase I and II clinical trials will ensue, and hopefully some of these agents will be found to be clinically promising. This approach, however, raises an interesting problem. If we examine the treatment of cancer by chemotherapy it is of course well known that single chemotherapeutic drugs are almost never used to treat the disease. Rather it is the *combination of different classes of drugs* which is the mainstay of chemotherapy protocols. With this analogy in mind, it would seem obvious that a similar 'combination'

anti-angiogenic therapy' might be necessary to detect successes in treating solid tumors by the strategy of inhibiting angiogenesis. Given the realities of how phase I clinical trials are conducted, and the issues concerning the commercialization and patents of new drugs, it may be extremely difficult to establish the right combination of circumstances required to detect optimal anti-angiogenic effects in patients with solid tumors. Thus combination antiangiogenic therapy will have to be explored primarily by basic researchers using appropriate animal models. It would seem worthwhile to consider seriously this type of therapeutic approach given the fact that there are many steps in the angiogenic process which appear susceptible to therapeutic attack. For example, endothelial cell invasion of surrounding basement membranes, adhesion to the subendothelial matrix, migration into the stroma and proliferation may all represent potential targets for anti-angiogenic drugs. Thus optimal antivascular therapy will likely include certain anti-integrin antibodies (e.g. anti- $\alpha v\beta 3$)⁵³ to inhibit endothelial cell invasion and/or adhesion in combination with other agents which, for example, inhibit endothelial cell proliferation, such as antibodies to VEGF/VPF or bFGF receptor tyrosine kinases.

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References

- Folkman J. Tumor angiogenesis. Adv Cancer Res 1985;
 43: 175-203.
- Dvorak HF, Nagy JA, Dvorak AM. Structure of solid tumors and their vasculature: implications for therapy with monoclonal antibodies. *Cancer Cells* 1991; 3: 77– 85.
- Rak JW, Hegmann EJ, Kerbel RS. The role of angiogenesis in tumor progression and metastasis. In: Heppner GH, ed. Advances in molecular and cell biology. Greenwich, CT: JAI Press 1993: 205-51.
- Blood CH, Zetter BR. Tumor interactions with the vasculature: angiogenesis and tumor metastasis. Bioch Biophys Acta 1990; 1032: 89–118.
- Bouch N. Tumor angiogenesis: the role of oncogenes and tumor suppressor genes. Cancer Cells 1990; 2: 179– 85.

- Paweletz N, Knierim M. Tumor related angiogenesis. In: CRC critical reviews in oncology/bematology. Orlando, FL: Academic Press 1989: 197–242.
- 7. Folkman J, Klagsbrun M. Angiogenic factors. *Science* 1987; **235**: 442–7.
- 8. Jain RK. Vascular and interstitial barriers to delivery of therapeutic agents in tumors. *Cancer Metastasis Rev* 1990; 9: 253-66.
- Rak JW, Hegmann EJ, Lu C, et al. Progressive loss of sensitivity to endothelium-derived growth inhibitors expressed by human melanoma cells during disease progression. J Cell Physiol 1994; 159: 245–55.
- 10. Hart IR. Easty D. Identification of genes controlling metastatic behaviour. *Br J Cancer* 1991; **63**: 9–12.
- 11. Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990; **61**: 759–67.
- 12. Nowell PC. The clonal evolution of tumor cell populations. *Science* 1976; **194**: 23–8.
- Khokha R, Waterhouse P, Yagel S, et al. Antisense RNAinduced reduction in murine TIMP levels confers oncogenicity on Swiss 3T3 cells. Science 1989; 243: 947–50.
- 14. Schipper JH, Frixen UH, Behrens J, et al. E-cadherin expression in squamous cell carcinomas of head and neck: inverse correlation with tumor differentiation and lymph node metastasis. Cancer Res 1991; 51: 6328–37.
- Eghbali B, Kessler JA, Reid LM, et al. Involvement of gap junctions in tumorigenesis: transfection of tumor cells with connexin 32 cDNA retards growth in vivo. Proc Natl Acad Sci USA 1991; 88: 10701-5.
- Good DJ, Polverini PJ, Rastinejad F, et al. A tumor suppressor-dependent inhibitor of angiogenesis is immunologically and functionally indistinguishable from a fragment of thrombospondin. Proc Natl Acad Sci USA 1990; 87: 6624–8.
- Moroco JR, Solt DB, Polverini PJ. Sequential loss of suppressor genes for three specific functions during in vivo carcinogenesis. Lab Invest 1990; 63: 298–306.
- Mahoney PA, Weber U, Onofrechuk P, et al. The fat tumor suppressor gene in *Drosophila* encodes a novel member of the cadherin gene superfamily. *Cell* 1991; 67: 853–68.
- 19. Dameron KM, Volpert OV, Tainsky MA, et al. p53 controls the switch to an angiogenic phenotype in fibroblasts by regulating thrombospondin. Science 1994; in press.
- Donehower LA, Harvey M, Slagle BL, et al. Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. Nature 1992; 356: 215–21.
- 21. Muller WJ. Expression of activated oncogenes in the murine mammary gland: transgenic models for human breast cancer. *Cancer Metastasis Rev* 1991; **10**: 217–27.
- 22. Kandel J, Bossy-Wetzel E, Radvanyi F, *et al.* Neovascularization is associated with a switch to the export of bFGF in the multistep development of fibrosarcoma. *Cell* 1991; **66**: 1095–104.
- Folkman J, Watson K, Ingber D, et al. Induction of angiogenesis during transition from hyperplasia to neoplasia. Nature 1989; 339: 58-61.
- 24. Scanlan MJ, Raj KBM, Calvo B, et al. Molecular cloning of fibroblast activation protein α, a member of the serine protease family selectively expressed in stromal fibroblasts of epithelial cancers. Proc Natl Acad Sci USA 1994; 91: 5457-661.

- 25. Cullen KJ, Smith SH, Hill S, et al. Growth factor messenger RNA expression by human breast fibroblasts from benign and malignant lesions. Cancer Res 1991; **51**: 4978–85.
- Basset P, Belloq JP, Wolf C, et al. A novel metalloproteinase gene specifically expressed in stromal cells of breast carcinomas. Nature 1990; 348: 699–704.
- Pyke C, Ralkiaer E, Tryggvason K, et al. Messenger RNA for two type IV collagenases is located in stromal cells in human colon cancer. Am J Pathol 1993; 142: 359–65.
- Pyke C, Kristensen P, Ralfkiaer E, et al. The plasminogen activation system in human colon cancer: messenger RNA for the inhibitor PAI-I is located in endothelial cells in the tumor stroma. Cancer Res 1991; 51: 4067–71
- 29. Folkman J. What is the evidence that tumors are angiogenesis-dependent? J Natl Cancer Inst 1990; 82: 4-6.
- Foulds L. Neoplastic development. London: Academic Press 1969.
- Nowell PC. Mechanisms of tumor progression. Cancer Res 1986; 46: 2203–7.
- Kerbel RS. Growth dominance of the metastatic cancer cell: cellular and molecular aspects. *Adv Cancer Res* 1990; 55: 87–132.
- 33. Bell CW, Pathak S, Frost P. Unknown primary tumors: establishment of cell lines, identification of chromosomal abnormalities, and implications for a second type of tumor progression. *Cancer Res* 1989; 49: 4311–15.
- 34. Brown K, Buchmann A, Balmain A. Carcinogen-induced mutations in the mouse c-Ha-*ras* gene provide evidence of multiple pathways of tumor progression. *Proc Natl Acad Sci USA* 1990; **87**: 538–42.
- van Meyel DJ, Ramsay DA, Keeney M, et al. p53 mutation, expression, and DNA ploidy in evolving gliomas: evidence for two pathways of progression. J Natl Cancer Inst 1994; 86: 1011-7.
- Rabbitts TH. Translocation, master genes, and differences between the origins of acute and chronic leukemias. Cell 1991; 67: 641–4.
- 37. Sidransky D, Mikkelsen T, Schwechheimer K, et al. Clonal expansion of p53 mutant cells is associated with brain tumour progression. *Nature* 1992; **355**: 846.
- 38. Parmiter AH, Nowell PC. Cytogenetics of melanocytic tumors. *J Invest Dermatol* 1993; **100**: 254s–8s.
- Woodruff MFA. Tumor clonality and its biological significance. Adv Cancer Res 1988; 50: 197-230.
- Fidler IJ. Critical factors in the biology of human cancer metastasis: twenty-eighth GHA Clowes memorial award lecture. Cancer Res 1990; 50: 6130–8.
- 41. Weiss L. Metastatic inefficiency. *Adv Cancer Res* 1990; **54**: 159–211.
- 42. Heppner GH. Tumor cell societies. J Natl Cancer Inst 1989; 81: 648-9.
- Miller BE, Miller FR, Heppner GH. Therapeutic perturbation of the tumor ecosystem in reconstructed heterogeneous mouse mammary tumors. *Cancer Res* 1989;
 3747–53.
- 44. Miller FR, Heppner GH. Cellular interactions in metastasis. *Cancer Metastasis Rev* 1990; 9: 21-34.
- Frankfurt OS, Seckinger D, Sugarbaker EV. Intercellular transfer of drug resistance. Cancer Res 1991; 51: 1190-5.
- Kobayashi H, Man S, Kapitain SJ, et al. Acquired multicellular-mediated resistance to alkylating agents in can-

- cer. Proc Natl Acad Sci USA 1993; 90: 3294-8.
- 47. Rak J. Possible role of tumour stem-end cell interaction in metastasis. *Med Hypotb* 1989; **29**: 17–9.
- 48. Raff M. Social controls on cell survival and cell death. *Nature* 1992; **356**: 397–400.
- 49. Folkman J. What is the role of angiogenesis in metastasis from cutaneous melanoma? Eur J Cancer Clin Oncol 1987; 23: 361-3.
- Polverini PJ, Cotran RS, Gimbrone MA, et al. Activated macrophages induce vascular proliferation. Nature 1977; 269: 804–6.
- 51. DiPietro LA, Polverini PJ. Angiogenic macrophages produce the angiogenic inhibitor thrombospondin 1. *Am J Pathol* 1993; **143**: 678.
- 52. Brooks PC, Clark RAF, Cheresh DA. Requirement of vascular integrin ανβ3 for angiogenesis. *Science* 1994; **264**: 569–72.
- 53. Ingber DE, Folkman J. How does extracellular matrix control capillary morphogenesis. *Cell* 1989; **58**: 803–5.
- 54. Liotta LA, Steeg PS, Stetler-Stevenson WG. Cancer metastasis and angiogenesis: an imbalance of positive and negative regulation. *Cell* 1991; **64**: 327–36.
- 55. Pepper MS, Belin D, Montesano R, et al. Transforming growth factor-beta 1 modulates basic fibroblast growth factor-induced proteolytic and angiogenic properties of endothelial cells in vitro. J Cell Biol 1990; 111: 743–55.
- Rooney P, Wang M, Kumar P, et al. Angiogenic oligosaccharides of hyaluronan enhance the production of collagens by endothelial cells. J Cell Science 1993; 105: 213-8.
- 57. Nguyen M, Strubel NA, Bischoff J. A role for sialyl Lewis-X/A gloycoconjugates in capillary morphogenesis. *Nature* 1993; **365**: 267–9.
- 58. Banerjee SD, Toole BP. Hyaluronan-binding protein in endothelial cell morphogenesis. *J Cell Biol* 1992; **119**: 643–52.
- Fotsis T, Zhang Y, Pepper MS, et al. The endogenous oestrogen metabolite 2-methoxyoestradiol inhibits angiogenesis and suppresses tumour growth. Nature 1994; 368: 237–55.
- Yamamoto T, Terada N, Nishizawa Y, et al. Angiostatic activities of medroxyprogesterone acetate and its analogues. Int J Cancer 1994; 56: 393–9.
- D'Amore PA, Braunhut SJ. Stimulatory and inhibitory factors in vascular growth control. *Endothelial Cells* 1988; II: 13-60.
- 62. Bickell R, Harris AL. Novel growth regulatory factors and tumor angiogenesis. *Eur J Cancer* 1991; 27: 781-5.
- Burri PH. Intussusceptive microvascular growth, a new mechanism of capillary network expansion. In: Angiogenesis: Int Symp, St Gallen, 1991; Abstract: 88.
- 64. Paku S, Lapis K. Morphological aspects of angiogenesis in experimental liver metastases. *Am J Pathol* 1993; **143**: 926–36.
- 65. Auerbach R, Auerbach W, Polakowski I. Assays for angiogenesis. *Pharmac Ther* 1991; **51**: 1-11.
- 66. Passaniti A, Taylor RM, Pili R, et al. Methods in laboratory investigation: A simple, quantitative method for assessing angiogenesis and antiangiogenic agents using reconstituted basement membrane, heparin, and fibroblast growth factor. Lab Invest 1992; 67: 519.
- Fajardo LF, Kwan HH, Kowalski J, et al. Dual role of tumor necrosis factor-α in angiogenesis. Am J Pathol

- 1992; **140**: 539.
- 68. Schweiger L, Neufeld G, Friedman J, et al. Capillary endothelial cells express basic fibroblast growth factor, a mitogen that promotes their own growth. *Nature* 1987; **325**: 257–9.
- Taraboletti G, Belotti D, Dejana E, et al. Endothelial cell migration and invasiveness are induced by a soluble factor produced by murine endothelioma cells transformed by polyoma virus middle t oncogene. Cancer Res 1993; 53: 3812–216.
- Dvorak HF. Tumors: wounds that do not heal. New Engl J Med 1986; 315: 1650-9.
- Senger DR, Van De Water L, Brown LF, et al. Vascular permeability factor (VPE, VEGF) in tumor biology. Cancer Metastasis Rev 1993; 12: 303–24.
- Dvorak HF, Sioussat TM, Brown LF, et al. Distribution of vascular permeability factor (Vascular Endothelial Growth Factor) in tumors: concentration in tumor blood vessels. J Exp Med 1991; 174: 1275–8.
- Plate KH, Breier G, Weich HA, et al. Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo. Nature 1992; 359: 845–8.
- Shweiki D, Itin A, Soffer D, et al. Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. Nature 1992; 359: 843-5.
- Jain RK. Barriers to drug delivery in solid tumors. Sci Am 1994; 58–65.
- 76. Heppner GH, Miller BE. Therapeutic implications of tumor heterogeneity. *Semin Oncol* 1989; **16**: 91-105.
- 77. Jouanneau J, Moens G, Bourgeois Y, et al. A minority of carcinoma cells producing acidic fibroblast growth factor induces a community effect for tumor progression. Proc Natl Acad Sci USA 1994; 91: 286–90.
- 78. Denijn M, Ruiter DJ. The possible role of angiogenesis in the metastatic potential of human melanoma. Clinicopathological aspects. *Melanoma Res* 1993; 3: 5–14.
- 79. Porschen R, Classen S, Piontek M, et al. Vascularization of carcinomas of the esophagus and its correlation with tumor proliferation. *Cancer Res* 1994; **54**: 587–91.
- 80. Koch AE, Polverini PJ, Kunkel SL, *et al.* Interleukin-8 as a macrophage-derived mediator of angiogenesis. *Science* 1992; **258**: 1798–801.
- 81. Weidner N, Carroll PR, Flax J, et al. Tumor angiogenesis correlates with metastasis in invasive prostate carcinoma. Am J Pathol 1993; 143: 401-9.
- 82. Guinebretiere J-M, Le Monique G, Gavoille A, et al. Angiogenesis and risk of breast cancer in women with fibrocystic disease. J Natl Cancer Inst 1994; 86: 635.
- Smith-McCune KK, Weidner N. Demonstration and characterization of the angiogenic properties of cervical dysplasia. *Cancer Res* 1994; 54: 800–4.
- 84. Weidner N, Semple JP, Welch WR, et al. Tumor angiogenesis and metastasis—correlation in invasive breast carcinoma. N Engl J Med 1991; 324: 1–8.
- 85. Folkman J. Angiogenesis and breast cancer. *J Clin On-* col 1994; **12**: 441-3.
- Srivastava A, Laidler P, Hughes LE, et al. Neovascularization in human cutaneous melanoma: a quantitative morphological and Doppler ultrasound study. Eur J Cancer Clin Oncol 1986; 22: 1205–9.
- Srivastava A, Laidler P, Davies RP, et al. The prognostic significance of tumor vascularity in intermediate-thick-

- ness (0.76–4.0 mm thick) skin melanoma. *Am J Pathol* 1988: **133**: 419–23.
- 88. Srivastava A, Hughes LE, Woodcock JP, *et al.* Vascularity in cutaneous melanoma detected by Doppler sonography and histology: correlation with tumour behaviour. *Br J Cancer* 1989; **59**: 89–91.
- 89. Barnhill RL, Levy MA. Regressing thin cutaneous malignant melanomas (≤1.0 mm) are associated with angiogenesis. *Am J Pathol* 1993; **143**: 99–104.
- Breslow A. Thickness, cross-sectional areas and depth of invasion in the prognosis of cutaneous melanoma. *Ann Surg* 1970; 172: 902–8.
- 91. Clark WH, Elder DE, Guerry D, et al. Model predicting survival in stage I melanoma based on tumor progression. J Natl Cancer Inst 1989; 81: 1893–1904.
- 92. Herlyn M. Human melanoma: development and progression. *Cancer Metastasis Rev* 1990; **9**: 101-12.
- Juhasz I, Albelda SM, Elder DE, et al. Growth and invasion of human melanomas in human skin grafted to immunodeficient mice. Am J Pathol 1993; 143: 528– 37
- 94. Kerbel RS. Expression of multi-cytokine resistance and multi-growth factor independence in advanced stage metastatic cancer: malignant melanoma as a paradigm. *Am J Pathol* 1992; **141**: 519–24.
- Clark WH, Elder DE, Guerry D, et al. A study of tumor progression: the precursor lesions of superficial spreading melanoma and nodular melanoma. Human Path 1984; 15: 1147-65.
- Reed JA, McNutt NS, Albino AP. Differential expression of basic fibroblast growth factor (bFGF) in melanocytic lesions demonstrated in situ hybridization. Am J Pathol 1994; 144: 329–36.
- 97. Mattei S, Colombo MP, Melani C, et al. Expression of cytokine/growth factors and their receptors in human melanoma and melanocytes. Int J Cancer 1994; 56: 853-7
- 98. Lu C, Rak JW, Kobayashi H, et al. Increased resistance to oncostatin M-induced growth inhibition of human melanoma cell lines derived from advanced-stage lesions. Cancer Res 1993; 53: 2708-11.
- Cornil I, Theodorescu D, Man S, et al. Fibroblast cell interactions with human melanoma cells affect tumor cell growth as a function of tumor progression. Proc Natl Acad Sci USA 1991; 88: 6028–32.
- 100. Lu C, Vickers MF, Kerbel RS. Interleukin-6: a fibroblast-derived growth inhibitor of human melanoma cells from early but not advanced stages of tumor progression. *Proc Natl Acad Sci USA* 1992; 89: 9215-9.
- 101. MacDougall J, Kerbel RS. Responsiveness of normal/dysplastic melanocytes and melanoma cells from different lesional stages of disease progression to the growth inhibitory effects of TGF-β. Mol Cell Different 1993; 1: 21–40.
- 102. Motro B, Itin A, Sachs L, et al. Pattern of interleukin 6 gene expression in vivo suggests a role for this cytokine in angiogenesis. Proc Natl Acad Sci USA 1990; 87: 3092–6.
- 103. Rodeck U, Melber K, Kath R, *et al.* Constitutive expression of multiple growth factor genes by melanoma cells but not normal melanocytes. *J Invest Dermatol* 1991; **97**: 20–6.
- 104. Gromet MA, Epstein WL, Blois MS. The regressing thin malignant melanoma—a distinctive lesion with meta-

- static potential. Cancer 1978; 42: 2282-92.
- 105. Graham CH, Rivers J, Kerbel RS, et al. Extent of vascularization as an independent prognostic indicator in thin (0.76 mm) malignant melanomas. Am J Pathol 1994, submitted.
- 106. Carnochan P, Briggs JC, Westbury G, et al. The vascularity of cutaneous melanoma: a quantitative histological study of lesions 0.85–125 mm thickness. Br J Cancer 1991; 64: 102–7.
- 107. Clark W. Tumor progression and the nature of cancer. *Br J Cancer* 1991; **64**: 631–44.
- 108. Sagebiel RW. Melanocytic nevi in histological association with primary cutaneous melanoma of superficial spreading and nodular types: effect of tumor thickness. *J Invest Dermatol* 1993; **100**: 3228–5S.
- 109. Tamm I, Cardinale I, Krueger J, et al. Interleukin 6 decreases cell-cell association and increases motility of ductal breast carcinoma cells. J Exp Med 1989; 170: 1649-69.
- 110. Zetter BR. Endothelial heterogeneity: influence of vessel size, organ localization, and species specificity on the properties of cultured endothelial cells. In: Ryan US, ed. *Endothelial Cells*, Boca Raton, FL: CRC Press 1988; 63.
- 111. Heimark RL, Schwartz SM. Endothelial morphogenesis. In: Simionescu N, Simionescu M, eds. *Endothelial cell biology in health and disease*. New York: Plenum Press 1988: 23–43.
- 112. Merwin JR, Newman W, Beall LD, et al. Vascular cells respond differentially to transforming growth factors beta1 and beta2 in vitro. Am J Pathol 1991; 138: 37-51.
- 113. Iruela-Arispe ML, Sage EH. Transforming growth factorβ promotes proliferation of endothelial cells that exhibit angiogenesis in vitro. In: The Molecular Biology of the Endothelial Cell. Keystone, CO: 1992: abstract 56.
- 114. Heffelfinger SC, Darlington G. SK-HEP-1: a model for angiogenesis. J Cell Biochem 1991; Supplement 15F: 250 (Abstract).
- 115. Auerbach R. Vascular endothelial cell differentiation: organ-specificity and selective affinities as the basis for developing anti-cancer strategies. *Int J Radiat Biol* 1991; 60: 1–10.
- 116. Sher-Taylor B, Bargatze R, Holzmann B, et al. Homing receptors and metastasis. Adv Cancer Res 1988; 51: 361.
- Nicolson GL. Organ specificity of tumor metastasis: role of preferential adhesion invasion and growth of malignant cells at specific secondary sites. *Cancer Metastasis Rev* 1988; 7: 143–88.
- 118. Hamada J., Cavanaugh PG, Miki K, et al. A paracrine migration-stimulating factor for metastatic tumor cells secreted by mouse hepatic sinusoidal endothelial cells: identification as complement component C3b. Cancer Res 1993; 53: 4418–23.
- Denekamp J. Vascular attack as a therapeutic strategy for cancer. Cancer Metastasis Rev 1990; 9: 267–82.
- 120. Clarke MSF, Kiff RS, Kumar S, et al. The identification of proliferation-related proteins in human endothelial cells as a possible target in tumour therapy. Int J Radiat Biol 1991; 60: 17–23.
- 121. Hagemeier H-H, Vollmer E, Goerdt S, et al. Monoclonal antibody reacting with endothelial cells budding vessels in tumors and inflammatory tissues, and nonreactive with normal adult tissues. Int J Cancer

- 1986; 38: 481-8.
- 122. Rettig WJ, Garin-Chesa P, Healey JH, et al. Identification of endosialin, a cell surface glycoprotein of vascular endothelial cells in human cancer. Proc Natl Acad Sci USA 1992; 89: 10832-6.
- 123. Koch AE, Nickoloff BJ, Holgersson I, *et al.* 4A11, a monoclonal antibody recognizing a novel antigen expressed on aberrant vascular endothelium. *Am J Pathol* 1994; **144**: 244–59.
- 124. Thorpe PE, Derbyshire EJ, King SW, et al. Targeting the vasculature of carcinomas and other solid tumors. *Proc Am Ass Cancer Res* 1994; 35: 379 (Abstract).
- 125. Wernert N, Raes M-B, Lassalle P, et al. c-ets1 protooncogene is a transcription factor expressed in endothelial cells during tumor vascularization and other forms of angiogenesis in humans. Am J Pathol 1992; 140: 119–27.
- 126. Partanen J, Armstrong E, Makela TP, et al. A novel endothelial cell surface receptor tyrosine kinase with extracellular epidermal growth factor homology domains. Mol Cell Biol 1992; 12: 1698–707.
- 127. Kaipainen A, Korhonen J, Pajusola K, et al. The related FLT4, FLT1, and KDR receptor tyrosine kinases show distinct expression patterns in human fetal endothelial cells. J Exp Med 1993; 178: 2077–88.
- 128. Yamaguchi TP, Dumont DJ, Conlon RA, et al. flk-1, an flt-related receptor tyrosine kinase is an early marker for endothelial cell precursors. Development 1993; 118: 489–98.
- 129. Dumont DJ, Yamaguchi TP, Conlon RA, et al. tek, a novel tyrosine kinase gene located on mouse chromosome 4, is expressed in endothelial cells and their presumptive precursors. Oncogene 1992; 7: 1471–80.
- 130. Runting AS, Stacker SA, Wilks AF. tie2, a putative protein tyrosine kinase from a new class of cell surface receptor. Growth Factors 1993; 9: 99-105.
- 131. Schnurch H, Risau W. Expression of *tie-*2, a member of a novel family of receptor tyrosine kinases, in the endothelial cell lineage. *Development* 1993; **119**: 957–68.
- 132. Millauer B, Wizigmann-Voos S, Schnurch H, et al. High affinity VEGF binding and developmental expression suggest Flk-1 as a major regulator of vasculogenesis and angiogenesis. Cell 1993; 72: 835–46.
- 133. Breier G, Albrecht U, Sterrer S, et al. Expression of vascular endothelial growth factor during embryonic angiogenesis and endothelial cell differentiation. Development 1992; 114: 521-32.
- 134. Quinn TP, Peters KG, de Vries C, et al. Fetal liver kinase 1 is a receptor for vascular endothelial growth factor and is selectively expressed in vascular endothelium. Proc Natl Acad Sci USA 1993; 90: 7533-7.
- 135. Kim KJ, Li B, Winer J, et al. Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumour growth in vivo. Nature 1993; 362: 841-4.
- 136. Millauer B, Shawver LK, Plate KH, et al. Glioblastoma growth inhibited in vivo by a dominant-negative Flk-1 mutant. Nature 1994; 367: 576-9.
- 137. Brown LF, Berse B, Jackman RW, et al. Increased expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in kidney and bladder carcinomas. Am J Pathol 1993; 143: 1255–62.
- 138. Plate KH, Breier G, Millauer B, et al. Up-regulation of

- vascular endothelial growth factor and its cognate receptors in a rat glioma model of tumor angiogenesis. *Cancer Res* 1993; **53**: 5822–7.
- 139. de Vries C, Escobedo JA, Ueno H, *et al.* The *fms*-like tyrosine kinase, a receptor for vascular endothelial growth factor. *Science* 1992; **255**: 989–91.
- 140. Greer P, Haigh J, Mbamalu G, Khoo W, Bernstein A, Pawson T. The Fps/fes protein-tyrosine kinase promotes angiogenesis in transgenic mice. *Mol Cel Biol* 1994; 14: 6755–63.
- 141. Ruta M, Howk R, Ricca G, et al. A novel protein tyrosine kinase gene whose expression is modulated during endothelial cell differentiation. Oncogene 1988; 3: 9-15.
- 142. Thorpe PE, Wallace PM, Knyba RE, et al. Targeting to proliferating vascular endothelium. Angiogenesis: Int Symp. St Gallen 1991: Abstract: 81.
- 143. Burrows FJ, Thorpe PE. Eradication of large solid tumors in mice with an immunotoxin directed against tumor vasculature. Proc Natl Acad Sci USA 1993; 90: 8996–9000.
- 144. Dvorak HF, Gresser I. Microvascular injury in pathogenesis of interferon-induced necrosis of subcutaneous tumors in mice. *J Natl Cancer Inst* 1989; 81: 497–502.
- 145. Folkman J, Ingher D. Inhibition of angiogenesis. *Semin Cancer Biol* 1993; **3**: 89–96.
- 146. Denekamp J. Angiogenesis, neovascular proliferation and vascular pathophysiology as targets for cancer therapy. Br J Radiol 1993; 66: 181–96.
- 147. Bicknell R, Harris AL. Anticancer strategies involving the vasculature: vascular targeting and the inhibition of angiogenesis. *Semin Cancer Biol* 1992; 3: 399–407.
- 148. Kerbel RS. Inhibition of tumor angiogenesis as a strategy to circumvent acquired resistance to anti-cancer therapeutic agents. *BioEssays* 1991; 13: 31-6.
- 149. Hori A, Sasada R, Matsutani E, et al. Suppression of solid tumor growth by immunoneutralizing monoclonal antibody against human basic fibroblast growth factor. Cancer Res 1991; 51: 6180-4.
- Wright PS, Cross-Doersen D, Miller JA, et al. Inhibition of angiogenesis in vitro and in ovo with an inhibitor of cellular protein kinases, MDL 27032. J Cell Physiol 1992; 152: 448-57.
- 151. Maier JAM, Voulalas P, Roeder D, et al. Extension of the life-span of human endothelial cells by an interleukin-1α antisense oligomer. Science 1990; 249: 1570–4.
- 152. Fox SB, Gatter KC, Bicknell R, et al. Relationship of endothelial cell proliferation to tumor vascularity in human breast cancer. Cancer Res 1993; 53: 4161-3.
- 153. Vartanian RK, Weidner N. Correlation of intratumoral endothelial cell proliferation with microvessel density (tumor angiogenesis) and tumor cell proliferation in breast carcinoma. Am J Pathol 1994; 144: 1188–94.
- Schipper H, Goh CR, Wang TL. Rethinking cancer: should we control rather than kill? Part 1. Can J Oncol 1993; 3: 207–16.
- 155. Rastinejad F, Polverini PJ, Bouck N. Regulation of the activity of a new inhibitor by angiogenesis by a cancer suppressor gene. *Cell* 1989; **56**: 345–55.
- 156. Heuer JG, von Bartheld CS, Kinoshitea Y, et al. Alternating phases of FGF receptor and NGF receptor expression in the developing chicken nervous system. Neuron 1990; 5: 283-96.

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- 157. Peters KG, Werner S, Chen G, et al. Two FGF receptor genes are differentially expressed in epithelial and mesenchymal tissues during limb formation and organogenesis in the mouse. Development 1992; 114: 233–43
- 158. Sato TN, Qin Y, Kozak CA, et al. tie-1 and tie-2 define another class of putative receptor tyrosine kinase genes expressed in early embryonic vascular system. Proc Natl Acad Sci USA 1993; 90: 9355-8.
- 159. Bilington DC. Angiogenesis and its inhibition: potential new therapies in oncology and non-neoplastic diseases. *Drug Design and Discovery* 1991; **8**: 3–35.
- 160. O'Reilly M, Holmgren L, Shing Y, Chen C, Rosenthal AR, Moses M, Lane WS, Cao Y, Sage EH, Folkman J. Angiostatin: a novel angiogenesis inhibitor that medtiates the suppression of metastases by Lewis lung carcinoma. *Cell* 1994; 79: 315–28.

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